

NbThermo: a new thermostability database for nanobodies

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Abstract

We present NbThermo—a first-in-class database that collects melting temperatures (T_m), amino acid sequences and several other categories of useful data for hundreds of nanobodies (Nbs), compiled from an extensive literature search. This so-far unique database currently contains up-to-date, manually curated data for 564 Nbs. It represents a contribution to efforts aimed at developing new algorithms for reliable T_m prediction to assist Nb engineering for a wide range of applications of these unique biomolecules. Nbs from the two most common source organisms—llama and camel—show similar distributions of melting temperatures. A first exploratory research that takes advantage of this large data collection evidences that understanding the structural bases of Nb thermostability is a complex task, since there are no apparent differences in sequence patterns between the frameworks of Nbs with lower and higher melting temperatures, indicating that the highly variable loops play a relevant role in defining Nb thermostability.

Database URL: <https://valdes-tresanco-ms.github.io/NbThermo>

Introduction

Heavy-chain antibodies (HCABs), commonly known as nanobodies (Nbs), have been gaining ground in the therapeutic and diagnostic areas since their discovery three decades ago. Common Nb applications include therapeutic uses in cancer and other diseases, *in vitro* and *in vivo* diagnosis, identification of molecular targets, drug delivery and as chaperone molecules in protein crystallography, among many others (1–4). Such versatility finds support in the unique structural characteristics of Nbs, such as the high thermal and reductive stabilities that allow their expression both extra- and intracellularly, their small size favoring tissue penetration, easy engineering allowing the generation of multimeric constructs, low immunogenicity due to their high similarity to the variable heavy domains of human antibodies and versatility in antigen recognition (5–7).

Intrinsic stability is of particular importance for the construction of synthetic Nb libraries (8), as well as for *in vitro* affinity maturation and the engineering of these molecules in general. Several studies have been performed aiming to understand both sequence and structural factors related to Nb stability, especially thermostability as measured through the melting temperature (T_m) (7, 9–14), which is the temperature at which approximately 50% of the protein molecules are unfolded. This property is closely related to other attributes

such as solubility and cell production. Thus, various studies have shown that mutation of one or several residues can increase T_m but drastically decrease yield (15, 16), while in other cases, it can affect the affinity for the antigen (15, 17–20).

The scarce availability of thermostability data for Nbs makes it difficult to generate algorithms to predict their T_m . Currently, several Nb databases have been reported, such as the ‘Integrated Nanobody Database for Immunoinformatics’ (21), SAbDab-nano (22), CoV-AbDab (23) and the ‘Institute Collection and Analysis of Nanobodies’ (iCAN) (24). These databases are focused mainly on sequences and crystallographic structures, although CoV-AbDab and iCAN also contain data on antigens and binding affinities. In this work, we present NbThermo—the first database that collects thermostability data for hundreds of Nb, together with amino acid sequences, 3D structures and other data that we considered important for studies that aim to understand the determinants of Nb stability.

Methods

Data collection

We started with an exhaustive search of the available literature in PubMed and Google Scholar, using the following

keywords: VHH, sdAbs, sybody, nanobody, HCAb, single domain antibody, antibody fragment, thermal stability and melting. With these keywords, we attempted to collect all the publications containing together any of the terms used for Nbs and thermostability data. Our search covered the time interval between 1993 (the year when HCABs in camelids were first reported) and 10 October 2022.

The following data were compiled for each database entry:

- (i) Name of the Nb.
- (ii) Reference DOI.
- (iii) Crystallographic structure (Protein Data Bank entry) when available.
- (iv) Melting temperature (T_m), subclassified according to the employed experimental method: nano differential scanning fluorimetry (nanoDSF), DSF with SYPRO Orange, differential scanning calorimetry (DSC), circular dichroism (CD) + refolding and ‘Other’ for methods not mentioned earlier.
- (v) Antigen, with the following fields: name, type and affinity.

- (vi) Cell production, subclassified according to the cellular location: periplasm, cytoplasm, and other for those without this information.
- (vii) Origin, subclassified into source for the organism (generic name, includes synthetic and unknown); type, according to whether the Nb is natural, fully synthetic or semi-synthetic; and obtaining method, which comprises immunization, naive, point mutation or other modification procedures.
- (viii) Sequence, with the following fields: raw sequence, sequence numbered using the Aho scheme (25), framework (Fw) and complementarity-determining regions (CDRs), labeled as follows: Fw1, CDR1, Fw2, CDR2, Fw3, CDR3, Fw4.

Annotations

Data found in text mode (e.g. Nb name, antigen name, DOI, sequence, etc.) were copied manually directly from the sources. Most of the sequences reported in the literature are found as images, and therefore, an online optical

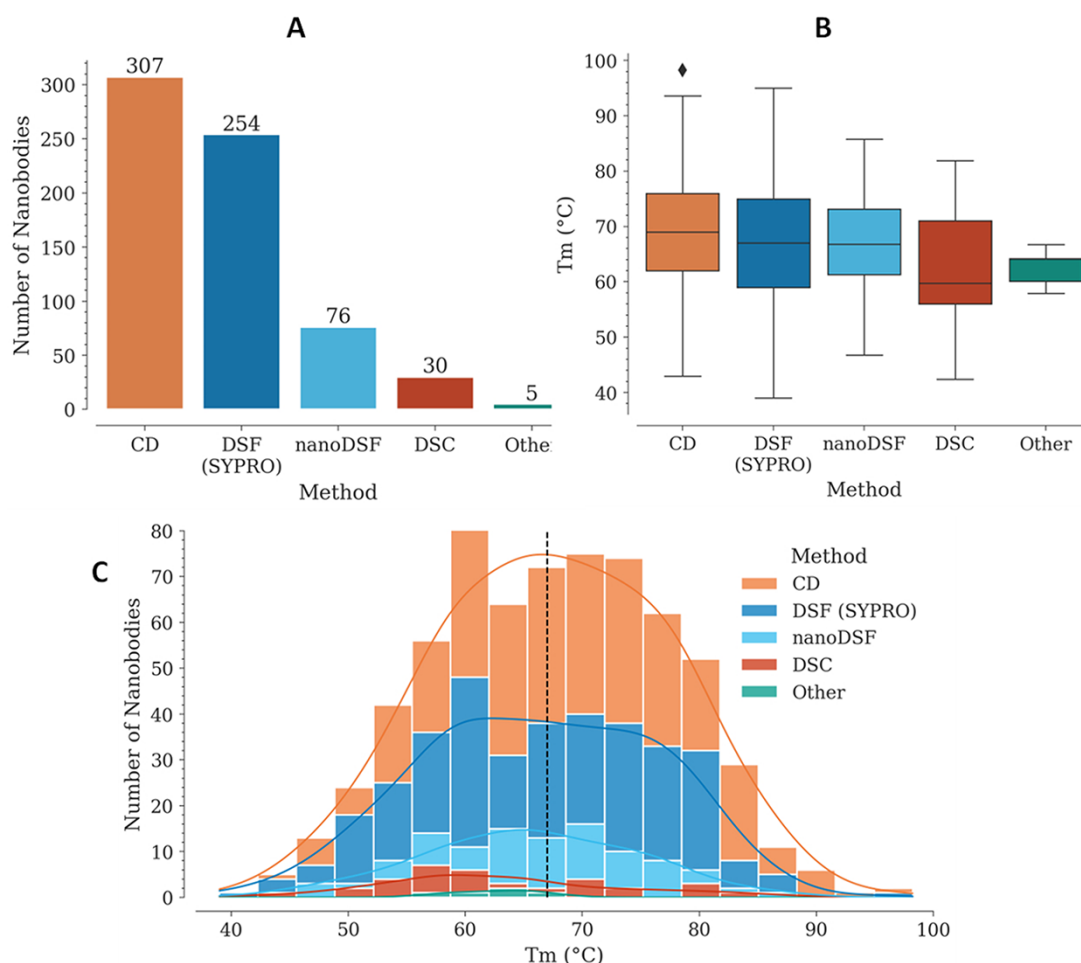


Figure 1. T_m data in the NbThermo database. (A) Distribution of T_m measurements by experimental methods: CD, DSF with SYPRO Orange, nanoDSF and DSC. (B) Distribution of T_m values as measured using different experimental methods. The box represents the interquartile range, the horizontal line within the box represents the median and the bottom and upper lines account for the minimum and maximum T_m , respectively. (C) Distribution of T_m values across the database, showing with stacked bars the use of the different experimental methods. The median value is highlighted by a vertical dashed line.

character recognition-to-text conversion tool (<https://www.ocr2edit.com/convert-to-txt>) was used, followed by a careful visual inspection of the obtained text. Each sequence, as well as mutant constructs whose sequences were not explicitly reported, was thoroughly checked. Likewise, antigen data such as affinity values (K_D —dissociation constant) as well as T_m and expression yields were manually annotated from tables in research articles and supplementary materials. K_D values were standardized in nanomolar (nM), while the expression yields were standardized in mg/l. The amino acid sequences were numbered according to the Aho scheme (25) using the ANARCI tool (26).

Implementation of NbThermo

A static website hosted on GitHub (<https://github.com/>) was created to implement and provide free access to NbThermo

(<https://valdes-tresanco-ms.github.io/NbThermo>). The frontend for this project was generated using Angular CLI version 13.2.4 (<https://angular.io/cli>).

Results

General overview

The literature search yielded ca. 1200 articles, which were all reviewed. As result, we collected data on 564 Nbs, found in 65 publications.

Melting temperature

For each Nb, T_m is reported with also the experimental method used for its measurement. In several cases, different values are reported for two or more experimental methods, for the same Nb. The method with the highest representation is CD, followed by DSF with SYPRO Orange, nanoDSE,

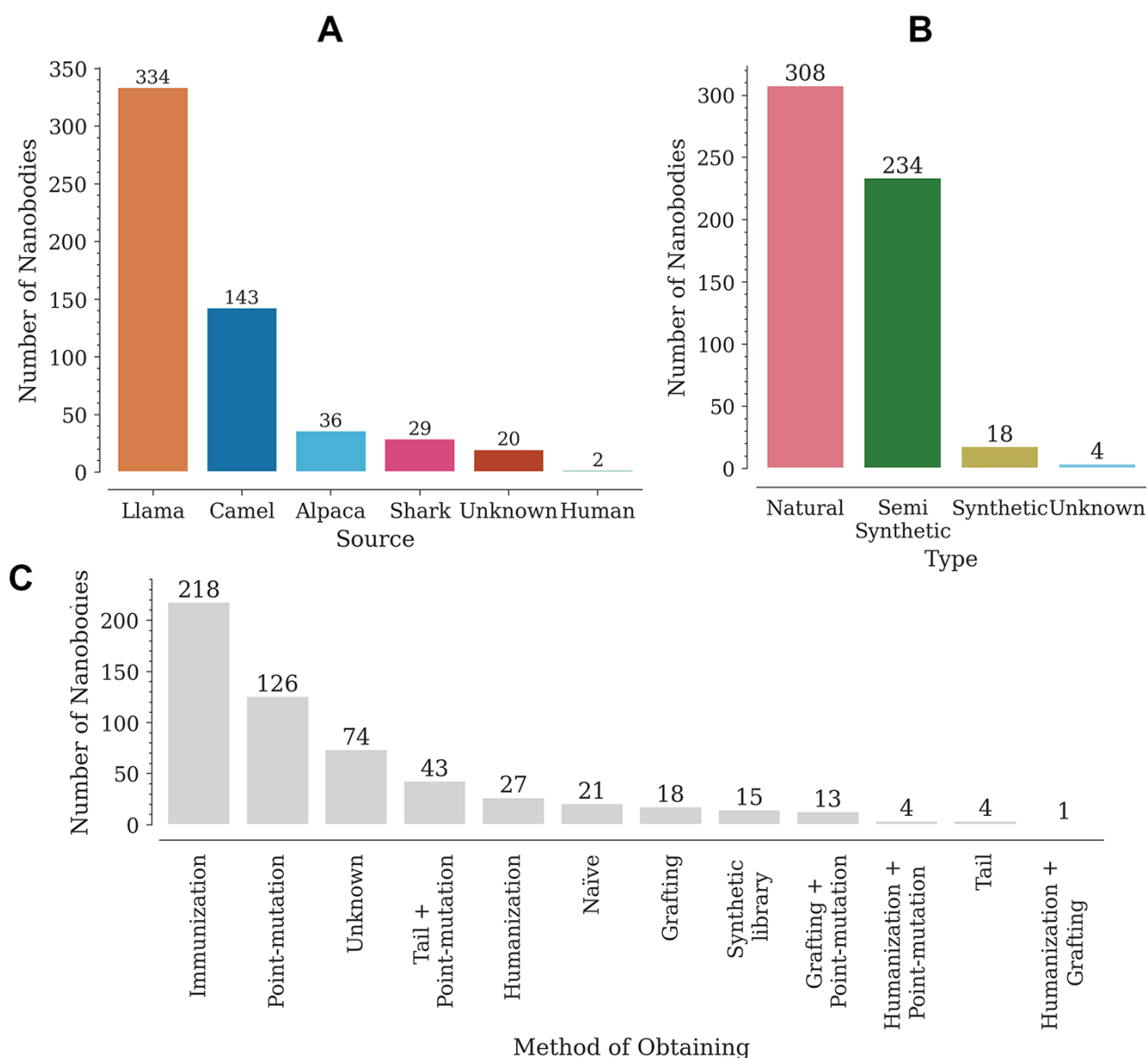


Figure 2. Distribution of Nbs in the database according to (A) source (organism), (B) type and (C) obtaining method.

DSC and a few cases where the experimental method is not reported (Figure 1A).

In most cases where T_m was measured using CD, the refolding ratio (secondary structure recovery) is also reported, evidencing that Nbs with high melting temperatures regain most of their secondary structure, with only a very few exceptions (data not shown). While T_m experiments using CD require specialized equipment, which limits its use, DSF with SYPRO Orange is more accessible since it can be performed with real-time PCR (RT-PCR) equipment and can cover a similarly broad T_m range (Figure 1B). Unlike with CD, Nb denaturalization with the SYPRO Orange reagent is irreversible, since it binds to the hydrophobic core of the structure. This is why using this method, it is not possible to measure the refolding ratio, but nonetheless, due to its higher accessibility, lower costs and ease of execution, its use for T_m determination is increasing.

Individually, the different experimental methods for T_m measurement show similar medians across the database: 69, 67, 67, 60 and 64°C, for CD, DSF (SYPRO), nanoDSF, DSC and ‘Other’, respectively (Figure 1B). Considering all the experimental methods, T_m is in the range from 39 to 98°C, with a median of 67.2°C (Figure 1C).

Nb origin and sequence

The database contains a wide variety of Nbs, coming mainly from llamas (302) and camels (175) (Figure 2A). The variable domains of the new antigen receptor of sharks, also included in the database, differ in sequence from camelid Nbs. Most of the reported Nbs are of natural origin, obtained from immune or naïve libraries (Figure 2B). Here, we have regarded as semi-synthetic all Nbs with at least one engineered modification, such as point mutations or tail additions (Figure 2B). It is worth noting that, although an N- or C-terminal addition does not modify the Nb sequence, it does have an effect on T_m . The Nbs in the database were obtained by multiple methods, with the most common being immunization and site-directed mutagenesis (point mutations) (Figure 2C). Animal immunization has been a widely used method to obtain high affinity binders, for which T_m is reported as stability data. Introducing

point mutations in some structures, on the other hand, aims at improving Nb–antigen affinity (*in vitro* affinity maturation) and/or improving other properties such as thermostability, solubility or cell production (7, 10, 12, 15, 16).

Manual annotation and revision of the amino acid sequences were undoubtedly the most complex task in the construction of this database. All the original sequences, including those with various tails such as His, Myc and Hop tags, were annotated. In addition, the Nb sequences were numbered according to the Aho scheme (25), which we selected among others mainly because it distributes the amino acids symmetrically around the gaps in the CDRs, thus piling up in the alignment those amino acids that are structurally close. This is useful especially for phylogenetic analyses of positional amino acid conservation/variation. Other numbering schemes will be added in the future.

For each sequence, we also annotated the lengths of its three CDRs. While CDR1 and CDR2 do not show important length variations (data not shown), CDR3 lengths span a wide range, from 3 to 26 amino acids, with lengths 15, 16 and 17 being the most represented in the database (Figure 3), similarly as reported in the recently published ‘Integrated nanobody database for immunoinformatics’ (21).

Other annotations

NbThermo includes also data related to Nb engineering. While mutations in the Fw region would rather not affect the affinity, mutations generated in the *in vitro* affinity maturation process may affect thermostability. We then included annotations on reported antigens (mostly of protein nature) and their affinities. Furthermore, some mutations can affect cell production, and therefore, we also included these data when available.

Implementation as website

NbThermo is freely available on its website (<https://valdes-tresanco-ms.github.io/NbThermo>) and in the repository (<https://github.com/Valdes-Tresanco-MS/NbThermo>). The graphical interface contains two panels (Figure 4): the left panel shows annotation filters (left) and the right panel

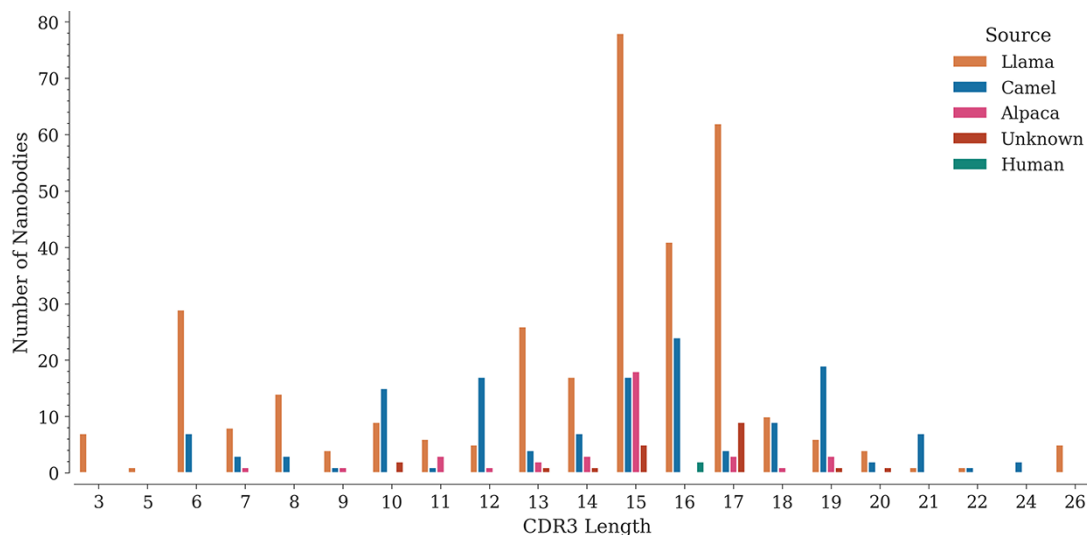


Figure 3. Distribution of CDR3 lengths in the database by source organism.

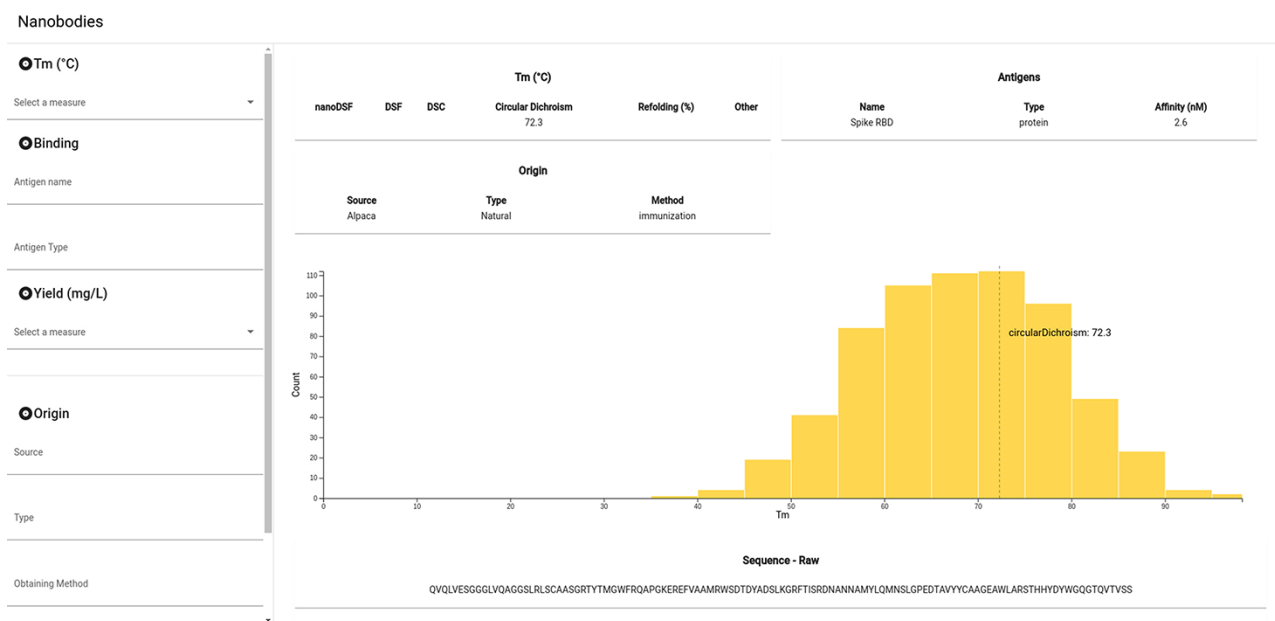


Figure 4. Snapshot of the NbThermo database interface. The left panel contains the annotation filters, while the right panel displays the selection list and the annotated information corresponding to the selected Nbs.

shows the selection list and the annotated information, where the information corresponding to the selected Nbs is given. The filter panel allows the user to explore on the fly amino acid sequences, T_m ranges, experimental methods, antigens and any other field in the database.

For each selected Nb, the data registered in the database are shown in the form of cards. In the same way, the T_m for the selected entry is highlighted in the T_m distribution graph of all the registered Nbs. Furthermore, users can access the original sequence, with its both sequential and Aho numbering schemes and information on the CDR and Fw regions. Finally, if available in the Protein Data Bank, the Nb three-dimensional structure can be inspected with an embedded molecular visualizer.

Exploring possible determinants of Nb thermostability

As an exploratory analysis of the data compiled in NbThermo, we set to investigate three questions of scientific and practical relevance, counting now with a richer source of data to support the analyses.

Q1—are there differences in the thermostability distribution for Nbs belonging to distinct species?

In a study by Kunz *et al.*, carried out with a dataset of 78 Nbs (camel—57, llama—17, alpaca—4, all of them of natural origin according to our classification), it was noted that llama Nbs tend to have a higher median T_m compared to camel Nbs (9). Performing a similar analysis in NbThermo yields the opposite result, although with a minor difference. Natural Nbs from camels show a higher median T_m (66 vs. 64°C for llama Nbs). However, if we include also Nbs of other origins, the overall medians for camel and llama Nbs are close to each other (camel T_m median = 68.5°C; llama T_m median = 68.0°C), followed by the median T_m values for alpacas

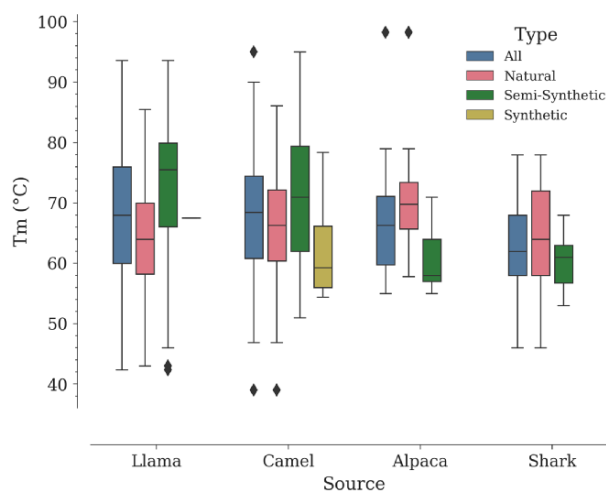


Figure 5. Distribution of T_m values by Nb: source and type.

and sharks (66 and 62°C, respectively) (Figure 5). Noteworthy, all organisms show a similar distribution around the overall database mean T_m value (67°C) (Figure S1). Since the difference between the median T_m of camels and llamas is small (2°C), the selection of one or the other species as a source of Nbs would be rather determined by other factors such as production costs.

Q2—is there a correlation between thermostability and CDR3 length?

We hypothesized that Nbs having longer CDR3, which are mostly bent over a Fw surface area that in classical antibodies interacts with the variable light domain, would show higher T_m values precisely because of the CDR3 packing against the Fw. However, only a poor correlation was obtained, which was slightly higher for camel Nbs (Figure S2). These results

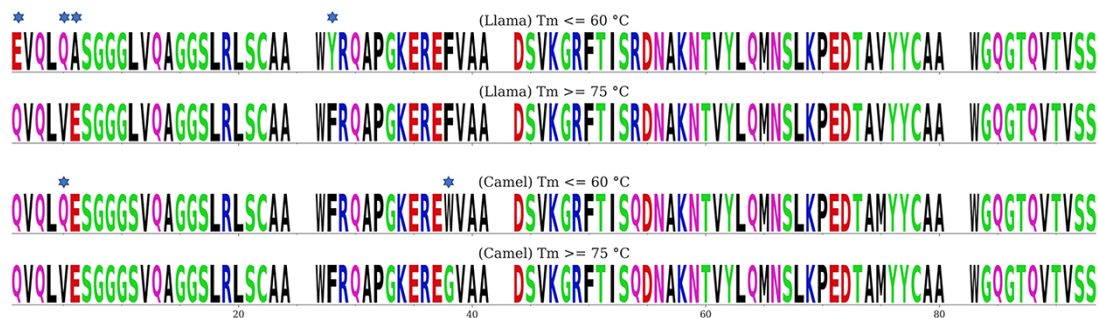


Figure 6. Llama and camel consensus Fw sequences for the low- and high- T_m groups. The white spaces correspond to the three CDRs. Sequence positions differing between the two groups of the same species are marked with an asterisk.

are similar to those obtained by Kunz *et al.* (7) using a smaller dataset.

Q3—are there different sequence patterns among the frameworks of Nbs with low and high thermostabilities?

Identifying sequence patterns in the Fw that would confer high thermostability to Nbs based on such Fws is of outmost importance for Nb engineering, particularly for the design of synthetic libraries. Here, we compared, both for camel and llama Nbs, two groups of Fw sequences having separated ranges of melting temperature: $T_m \leq 60^\circ\text{C}$ and $T_m \geq 75^\circ\text{C}$. For each group, we extracted the consensus Fw sequence (Figure 6).

Interestingly, only four differences in the consensus Fw sequences (E1Q, Q5V, A6E and Y28F) were found between the low- and high- T_m llama groups, while only two differences were found for the camel groups (Q5V and W38G). It has been shown that even a single mutation may cause a considerable T_m variation; however, these few differences in consensus sequence can hardly explain the T_m differences between the low- and high- T_m groups since the differing amino acids are present in both groups (Figure S3). For example, in camel Nbs, both tryptophan and glycine are found at position 38 in both the low- and high- T_m groups, although at different frequencies. Interestingly, comparison of the consensus sequences between the high- T_m groups in llama and camel shows that they differ by the same number of positions as the high- and low- T_m groups in llama (Table S1).

These findings indicate that the Fw structure alone does not define Nb thermostability, implying that the CDRs make an essential contribution. In particular, the interaction between CDR3 and the Fw may play a significant role, as found in a recent study by Kinoshita *et al.*, showing that mutations of amino acids at this interface can cause a considerable T_m decrease (27). Thus, the so-called ‘universality’ of a Fw, relevant for synthetic library construction, would then rely on the Fw adaptability to variable CDR3 characteristics, such as length, amino acid composition and conformation.

Noteworthy, when performing a similar sequence comparison for Nbs for which refolding data are available (the analysis was carried out only for llama Nbs since too little data were found for other camelid species), more variability is observed between the consensus Fw sequences of the Nb groups with low (< 50%) and high (> 75%) refolding ratios (Figure S4). Interestingly, most of the observed differences correspond to solvent-accessible positions. A moderate

correlation between T_m and refolding is observed ($r_p = 0.63$ and $r_s = 0.64$). However, it is difficult to draw meaningful conclusions because of the limited amount of data on folding recovery in the database.

Projections

The current implementation of NbThermo allows visualization and retrieval of stored data. Various filters can be applied to select sets of Nbs belonging to distinct categories or having melting temperatures within a desired range. Several features will be improved in the future, such as interactive search based on sequence identity, annotation of new entries by community contributions, artificial intelligence–based structural models and T_m prediction for new sequences.

Conclusions

Here, we presented NbThermo—a curated thermostability database of Nbs. The database contains up-to-date data for 564 Nbs, which were compiled from diverse sources and manually annotated to ensure maximum reliability. Our exploratory research, now using a much larger amount of data compared to previous studies, indicates that understanding the structural bases of Nb thermostability is a complex task since both the Fw and the CDR regions play significant roles and there are no evident differences in sequence patterns between the Fws of Nbs with lower and higher melting temperatures. We have put effort in compiling these data hoping to contribute to the development of new algorithms for reliable T_m prediction, as useful tools to assist Nb engineering for the wide range of applications of these unique biomolecules.

Supplementary material

Supplementary material is available at *Database* online.

Data availability

The NbThermo database is available at <https://valdes-tresanco-ms.github.io/NbThermo>.

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Conflict of interest

None declared.

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